# Melamine, Cyanuric Acid, Ammeline, and Ammelide Analysis by Gas Chromatography / Mass Spectrometry cat.# 33254

#### Melamine Analysis Kit (cat.# 33254) Includes:

······································	
GC Column	
Description	cat.#
Rxi-5Sil MS w/5m Integra-Guard Column (30m, 0.25mm ID, 0.25µm)	13623-124
Standards:	
Volume is 1mL/ampul. Concentration is 1,000µg/mL in diethylamine:water (20:80).	
Description	cat.#
Melamine Stock Standard	33247
Cyanuric Acid Stock Standard	33248
Ammelide Stock Standard	33249
Ammeline Stock Standard	33250
Benzoguanamine Internal Standard	33251
Melamine and Related Analogs Stock Standard	
(1mL ampul containing 1,000µg/mL each: ammelide, ammeline, cyanuric acid, and melamine)	33253
Derivatization Reagent:	
Description	cat.#
25g vial BSTFA w/ 1% TMCS	35607
Accessories:	
Description	cat.#
50mL empty centrifuge tube, 25-pk.	26227*
13mm, 0.45µm nylon syringe filter, 100-pk.	26147*

\*Kit contains a 5-pack each of tubes and filters. 25-pks (cat.# 26227) and 100-pks (cat.# 26147) sold separately.

#### Introduction

The recent discovery of melamine in infant formula and pet food has raised industry, government, and public concerns about the potential for more widespread contamination. Melamine is not a legal food additive; it is a nitrogen-rich industrial compound used for plastics, flame-resistant products, and cleaning agents. However, melamine and related byproducts have been illegally added to food products in order to falsely represent the amount of protein present, since protein level in many products is determined using nonspecific assays for nitrogen content (Figure 1). While nontoxic alone at low doses, melamine does pose significant health risks, including kidney failure, when consumed in combination with cyanuric acid.

In response to the increasing need for more rigorous melamine and cyanuric acid testing, the US Food and Drug Administration (FDA) has set three different commodity-based minimum reporting levels (MRLs):  $10\mu g/g$  for pet food,  $2.5\mu g/g$  for human food, and  $1\mu g/g$  for infant formula. The lower MRL level for infant formula was established because infants may rely on formula as their sole food source and since their renal systems are not yet fully developed.

#### **Method Overview**

The products and step-by-step instructions included in this kit are designed to provide a complete solution for the GC/MS analysis of melamine and related compounds. The extraction and derivatization method described here was adapted from the FDA method, *GC-MS Screen for the Presence of Melamine, Ammeline, Ammelide, and Cyanuric Acid.*<sup>1</sup> Please refer to the FDA method for quality control (QC) requirements, reporting criteria, and semi-quantitative calculations.

Note that the FDA method guidelines differ based on the MRL for the commodity being analyzed; both high and low concentrations of standards and matrix spikes may be analyzed in the same analytical set for  $10\mu g/g$  MRL commodities, but should be analyzed in separate sets for lower MRL commodities to reduce the potential for carryover contamination. For  $2.5\mu g/g$  and  $1\mu g/g$  MRL





commodities, first analyze derivatized samples with the low matrix spike and standards. Then, if target analytes are detected above the MRL and semi-quantitative results are desired, new aliquots may be derivatized and analyzed with the high standards (and a high matrix spike, if desired). Instructions are provided here for the preparation of both high and low levels of QC matrix spikes and quantification standards for each MRL (Tables I and II). Example data are shown in Figures 2-7.

Finally, care must be taken to monitor for matrix interferences, which may complicate data collection and interpretation, as well as contaminate instrumentation. Analyzing derivatization reagent blanks at appropriate intervals is recommended. It may be necessary to perform a sample clean-up procedure to help reduce matrix interferences.



### Procedure

## Sample and Standard Preparation

Prepare stock standard solutions at the appropriate levels for the commodity being analyzed according to Table I. Next, use these stock solutions to make the actual samples and standards for analysis according to Table II. Some matrices have an analyte protecting effect, causing an increased response compared to analysis with solvent-only standards.<sup>2,3,4</sup> To avoid this effect, use matrix-matched standards, instead of solvent-based standards, for 2.5µg/g and 1µg/g MRL commodities, as shown in Table I. Matrix-matched standards are prepared in extracted analyte-free control matrix instead of solvent. Both matrix-matched and solvent-based standards must be derivatized according to the procedure in this method.

High and low matrix spikes are prepared using representative control sample matrix according to Table II and then extracted and derivatized. Low matrix spikes are prepared at the MRL to demonstrate adequate instrument sensitivity; thus, the low spike must be observed in order to report negative samples as below the MRL. High spikes are optional and can be useful indicators of major problems with sample extraction or system performance. A method solvent blank (20mL of 10/40/50 DEA/H<sub>2</sub>O/ACN) should also be extracted to ensure there is no contamination from the reagents, nylon filter, or instrument system.

	10µg/g MRL (Pet Food)	2.5µg/g MRL (Human Food)	1µg∕g MRL (Infant Formula)
Mixed Standard	Target: 100µg/mL	Target: 50µg/mL	Target: 10µg/mL
Stock Standard	1mL stock standard diluted	500µL stock standard diluted	100µL stock standard diluted
1,000µg/mL	to 10mL with 10/40/50	to 10mL with 10/40/50	to 10mL with 10/40/50
(cat.# 33253)	DEA/H2O/ACN*	DEA/H <sub>2</sub> O/ACN	DEA/H <sub>2</sub> O/ACN
Internal Standard	Target: 10µg/mL	Target: 1µg/mL	Target: 0.5µg/mL
Benzoguanamine	$100\mu$ L benzoguanamine diluted	$10\mu$ L benzoguanamine	$5\mu$ L benzoguanamine diluted
1,000µg/mL	to 10mL with pyridine	diluted to 10mL with pyridine	to 10mL with pyridine
High Standard	Target: 10µg/mL	Target: 1µg/mL	Target: 0.25µg/mL
Prep Solution**	100µL Mixed Standard diluted	20µL Mixed Standard diluted	25µL Mixed Standard diluted to
	to 1mL with 10/40/50 DEA/H2O/ACN	to 1mL with control matrix extract	1mL with control matrix extract
Low Standard	Target: 1µg/mL	Target: 0.25µg/mL	Target: 0.1µg/mL
Prep Solution**	10µL Mixed Standard diluted	5µL Mixed Standard diluted	10µL Mixed Standard diluted to
	to 1mL with 10/40/50 DEA/H <sub>2</sub> O/ACN	to 1mL with control matrix extract	1mL with control matrix extract

	10µg/g MRL (Pet Food)	2.5µg/g MRL (Human Food)	גµg/g MRL (Infant Formula)
	Target: $1\mu g/mL$	Target: 0.1µg/mL	Target: 0.25µg/mL
High Standard*	Aliquot 50µL of High Standard	Aliquot 50µL of High Standard	Aliquot 50µL of High Standard
	Prep Solution and derivatize.	Prep Solution and derivatize.	Prep Solution and derivatize.
	Target: 0.1µg/mL	Target: 1µg/mL	<u>Target: 0.01µg/mL</u>
Low Standard	Aliquot 50µL of Low Standard	Standard Prep Solution	Prep Solution and derivatize.
	and derivatize.	and derivatize.	
	Target: 50µg/g	Target: 5µg/g	Target: 2.5µg/g
High Matrix	250µL Mixed Standard in 0.5g	50 $\mu$ L Mixed Standard in 0.5g	125 $\mu$ L Mixed Standard in 0.5g
Spike*	matrix. Extract and derivatize.	matrix. Extract and derivatize.	matrix. Extract and derivatize
	Target: 10µg/g	Target: 2.5µg/g	Target: 1µg/g
Low Matrix Spike	50 $\mu$ L Mixed Standard in 0.5g	$25\mu$ L Mixed Standard in	50 $\mu$ L Mixed Standard in 0.5g
	matrix. Extract and derivatize.	0.5g matrix. Extract and derivatize.	matrix. Extract and derivatize.

#### **Matrix Extraction**

Use the following procedure to extract test samples, matrix blanks, and matrix spikes. (Extracts from matrix blanks can also be used to prepare matrix-matched standard prep solutions according to Table I.)

- Weigh out 0.5g of each test or control sample into a 50mL centrifuge tube (cat.# 26227).
- Prepare high and low matrix spikes using control matrix according to the instructions in Table I (omit this step for matrix control and test samples).
- Add 20mL of 10/40/50 DEA/H<sub>2</sub>O/ACN solution.
- · Mix well to wet entire sample.
- Sonicate for 30 minutes.
- Centrifuge for 10 minutes to pellet.
- Filter 2mL portion with 0.45 $\mu$ m nylon filter (cat.# 26147).

Note: A 2mL portion is filtered to allow a second aliquot to be derivatized, if necessary.

REDUCK © 2009 Restek Corporation

### Derivatization

The derivatization procedure is necessary to allow melamine and related analogs to become volatile in the injection port of the gas chromatograph. Derivatize prepared samples and standards according to the following procedure.

- Transfer the following volumes to individual vials:
- Sample supernatants: 200µL.
- Matrix spike supernatants: 200µL.
- Matrix-matched standards (high and low): 200µL.
- Solvent-based standards (high and low): 50µL.
- Evaporate to dryness at 70°C with inert gas. Verify that samples are dry; remaining water can disrupt the derivatization reaction.
- Add 200µL pyridine.
- Add 200µL BSTFA with 1% TMCS derivatizing reagent.
- Add 100µL of benzoguanamine internal standard.
- · Shake well or vortex for 10 minutes.
- Incubate at 70°C for 45 minutes.
- Inject into the GC/MS system.

#### Analytical Set

Standards should be injected both before and after the sample set for semi-quantitative analysis; however, the use of the high and low standards differs by the MRL. For 10µg/g MRL commodities, both low and high standards can be run in a single analytical set. For 2.5µg/g and 1µg/g MRL commodities, first analyze the samples with the low standards (do not inject the high standard). Then, if target analytes are detected above the MRL, new aliquots may be derivatized and analyzed separately with the high standards in a new analytical set. Low matrix spikes serve to establish detectability at the MRL and must be detected in order to report negative sample results as less than the MRL. The analysis of derivatization reagent blanks at appropriate intervals is recommended to detect carryover.

#### **GC/MS** Parameters Col

Column:	Rxi <sup>®</sup> -5Sil MS, 30m x 0.25mm ID x 0.25µm with 5m Integra-Guard <sup>™</sup> (cat.# 13623-124)
Instrument:	Shimadzu QP 2010 Plus
Injection mode:	splitless, 1 minute sampling time
Injection liner:	splitless, 3.5mm Gooseneck Splitless Line with wool (cat.# 2228-200.1)
Injection volume:	1µL
Inlet temp.:	280°C
Carrier gas:	helium
Linear velocity:	37cm/sec., constant linear velocity
Oven temp.:	75°C to 320°C @ 15°C/min.
-	(hold 4 min.)
Detector:	MS
Transfer line temp.:	290°C
Scan range:	50-450 amu
Ionization:	EI, ion source at 190°C
Filament delay:	8 minutes
Mode:	scan or SIM

#### Other liner choices by instrument:

Agilent Gooseneck Splitless 4mm with wool (cat.# 22405) PerkinElmer Auto SYS Splitless with wool (cat.# 20829) Shimadzu Gooseneck Splitless with wool for 17A, 2010, 2014 (cat.# 22286-200.1) Shimadzu Splitless with wool for 14A (cat.# 20863-200.1) Thermo 4000/5000/6000 Splitless with wool (cat.# 20814-200.1) Thermo TRACE, 8000, 8000 TOP, Focus SSL Splitless with wool (cat.# 20942-200.1) Varian 1075/1077 4mm Splitless with wool (cat.# 20904-200.1) Varian 1177 Gooseneck Splitless with wool (cat.# 21896-200.1) Varian 1078/1079 Splitless with wool (cat.# 21711-200.1)

#### MS Conditions (SIM mode)

Compound	Retention	Target	Reference	Reference	Reference
	Time (min)	Ions	Ions	Ions	Ions
Cyanuric Acid	10.23	345	330	346	347
		(100)*	(36)	(30)	(15)
Ammelide	11.07	344	329	345	330
		(100)	(58)	(30)	(16)
Ammeline	11.76	328	343	329	344
		(100)	(79)	(29)	(24)
Melamine	12.31	327	342	328	343
		(100)	(53)	(30)	(17)
Benzoguanamine	14.54	316	331	332	330
		(100)	(68)	(20)	(9)

\*relative ion ratio

	Solvent Blank	Matrix Blank	Spike ( µg/g)	Spike ( µg/g)	Sample #	Sample #	Sample #	Sample #	Sample #
Actual weight (g)			•						
Fortification									
Add 20mL solvent (10:40:50 DEA:H20:ACN)									
Sonicate 30 min. start time: end time:									
Centrifuge 10 min. start time: end time:									
<b>Filter</b> supernatant (0.45μm nylon syringe filter)									
<b>Evaporate</b> filtered extract aliquot* to dryness (70°C & nitrogen gas) start time: end time:									
<b>Add</b> 200µL pyridine									
Add 200µL BSTFA with 1% TMCS									
<b>Add</b> 100µL benzoguanamine**									
Shake well (10 min.)									
Incubate at 70°C for 45 min. start time: end time:									
Inject ( $\mu$ L)									
Injection concentration (spikes)									
Aliquot volume varies t <b>ype</b> olvent-based standard datrix-based standard amples & spikes	y supernatant type. Aliquot 50µL 200µL 200µL		** The recommended in standard) is based on th MRL 10µg/g (pet food) 2.5µg/g (human food) 1.0µg/g (infant formula)	iection concentration of t ne MRL of the commodity Benzoguanamine injectić 0.2µg/mL 0.1µg/mL	enzoguanamine (internal being analyzed. <b>on conc.</b>		Method: Analyst: Date:		

# Melamine Analysis Fortification and Extraction Sheet (QC Included) © 2009 Restek Corporation



















#### References

- 1 US Food and Drug Administration, October 2008, GC-MS Screen for the Presence of Melamine, Ammeline, Ammeline, and Cyanuric Acid, Laboratory Information Bulletin No. 4423, http://www.cfsan.fda.gov/~frf/lib4423.html.
- 2 C.F. Poole, J. Chromatogr. A 1158 (2007) 241.
- 3 T. Cajka, K. Maštovská, S.J. Lehotay, J. Hajšlová, J. Sep. Sci. 28 (2005) 1048.
- 4 K. Maštovská, S.J. Lehotay, M. Anastassiades, Anal. Chem. 77 (2005) 8129.

#### Call Technical Service at 800-356-1688 or 814-353-1300, ext. 4 (or your Restek representative) if you have any questions about this product or any other Restek product.





#415-01 [001] Rest Rev. date: 02/09 Thar

Restek U.S. • 110 Benner Circle • Bellefonte, PA 16823 phone: 814-353-1300 • 800-356-1688 • fax: 814-353-1309 • www.restek.com Restek France • phone: +33 (0)1 60 78 32 10 • fax: +33 (0)1 60 78 70 90 • e-mail: restek@restekfrance.fr Restek GmbH • phone: +49 (0)6172 2797 0 • fax: +49 (0)6172 2797 77 • e-mail: info@restekgmbh.de Restek Ireland • phone: +44 (0)2890 814576 • fax: +44 (0)2890 814576 • e-mail: restekeurope@aol.com Thames Restek U.K. LTD • phone: +44 (0)1494 563377 • fax: +44 (0)1494 564990 • e-mail: sales@thamesrestek.co.uk